

Positron Emission Tomography Studies of Potential Mechanisms Underlying Phencyclidine-Induced Alterations in Striatal Dopamine

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Positron emission tomography (PET), in combination with ¹¹C-raclopride, was used to examine the effects of phencyclidine (PCP) on dopamine (DA) in the primate striatum. In addition, we explored the hypotheses that GABAergic pathways as well as molecular targets beyond the *N*-methyl-D-aspartate (NMDA) receptor complex (ie dopamine transporter proteins, DAT) contribute to PCP's effects. In the first series of experiments, ¹¹C-raclopride was administered at baseline and 30 min following intravenous PCP administration. In the second series of studies, γ -vinyl GABA (GVG) was used to assess whether enhanced GABAergic tone altered NMDA antagonist-induced changes in DA. Animals received an initial PET scan followed by pretreatment with GVG (300 mg/kg), then PCP 30 min prior to a second scan. Finally, we explored the possible contributions of DAT blockade to PCP-induced increases in DA. By examining ¹¹C-cocaine binding a paradigm in which PCP was coadministered with the radiotracer, we assessed the direct competition between these two compounds for the DAT. At 0.1, 0.5, and 1.0 mg/kg, PCP decreased ¹¹C-raclopride binding by 2.1, 14.9 ± 2.2 and $8.18 \pm 1.1\%$, respectively. These effects were completely attenuated by GVG ($3.38 \pm 3.1\%$ decrease in ¹¹C-raclopride binding). Finally, PCP (0.5 mg/kg) decreased ¹¹C-cocaine binding by $25.5 \pm 4.3\%$, while at 1.0 mg/kg this decrease was 13.5%, consistent with a competitive interaction at the DAT. These results suggest that PCP may be exerting some direct effects through the DAT and that GABA partially modulates NMDA-antagonist-induced increases in striatal DA.

Neuropsychopharmacology (2003) **28**, 2192–2198, advance online publication, 23 July 2003; doi:10.1038/sj.npp.1300258

Keywords: positron emission tomography (PET); schizophrenia; GABA; phencyclidine; NMDA antagonist; dopamine; vigabatrin

INTRODUCTION

Phencyclidine (PCP), an *N*-methyl-D-aspartate (NMDA) receptor antagonist, was developed more than four decades ago as an intravenous anesthetic. Its use was discontinued when it became apparent that it was addictive and frequently produced psychological dependence, compulsive drug-seeking behavior, and profound psychotic episodes. According to the National Household Survey on Drug Abuse (NHSDA), 3.2% of the population aged 12 years and older have used PCP at least once, and the abuse of its cogener and popular 'club drug', ketamine, has increased dramatically since 1999. While PCP and ketamine are readily self-administered by animals (Rodefer and Carroll, 1999), the role and extent of reward-related dopaminergic processes in these behaviors remain to be established. This becomes even more critical with respect to the similarity

between PCP-induced psychoses and schizophrenic syndromes that are partially ameliorated by dopamine (DA) antagonist drugs.

Rapid increases in striatal DA appear to represent an integral component of the self-reported 'high', associated with psychostimulants (Volkow *et al*, 1999). These increases may be achieved directly through interactions with DA releasing or reuptake mechanisms, or indirectly through interactions with other functionally linked neurotransmitter systems. For example, the initial molecular targets of most drugs of abuse, including the opiates, nicotine, and PCP are not dopaminergic. In fact, all these compounds exert their effects through multiple efferent pathways that originate from sites distal to the mesolimbic DA system. With respect to PCP and ketamine, the mechanisms that connect NMDA receptor blockade to striatal DA systems remain undetermined. Some evidence indicates that like the opiates, antagonism of NMDA receptors may prevent the excitation of inhibitory GABAergic neurons that normally modulate these pathways (Grunze *et al*, 1996; Li *et al*, 2002). This disinhibition may contribute to the reward-related DA response in the striatum and other mesolimbic and mesocortical brain regions, and may be one mechanism involved in NMDA antagonist-induced psychoses (Olney

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Received 25 February 2003; revised 22 May 2003; accepted 30 May 2003

Online publication: 5 June 2003 at <http://www.acnp.org/citations/Npp060503030084/default.pdf>

and Farber, 1995). Alternatively, PCP and/or ketamine may stimulate striatal DA through a direct interaction between glutamatergic and dopaminergic neurons (Adams and Moghaddam, 1998). Further complicating possible interactions with DA systems is PCP's relatively potent affinity for the dopamine transporter (DAT) site; an effect not shared by its congener, MK-801 (Johnson and Snell, 1985; Maurice *et al*, 1991). Nevertheless, since the affinity of PCP for the NMDA receptor is so much greater than its affinity for the DAT, it has been suggested that blockade of transporter proteins is unlikely to contribute significantly to either its behavioral or neurochemical response (Carlezon and Wise, 1996). Moreover, the involvement of DA systems in the rewarding or psychotomimetic properties of PCP has yet to be unequivocally established, and recent studies suggest that NMDA antagonists may exert their effects in the absence of any dopaminergic contribution (Adams *et al*, 2002; Kegeles *et al*, 2002).

Therefore, at the most fundamental level, it becomes critical to characterize the dopaminergic response to specific NMDA antagonists. In doing so, novel experimental strategies can begin to unravel the putative mechanisms involved in this response. Positron emission tomography (PET) provides the unique opportunity to observe changes in synaptic DA noninvasively by measuring changes in radiotracer binding. Our laboratory and others have used this technique to provide evidence that ketamine significantly increases synaptic DA transmission (Breier *et al*, 1998; Smith *et al*, 1998; Tsukada *et al*, 2000; Vollenweider *et al*, 2000). We have also successfully employed this approach to characterize functional interactions between cholinergic, GABAergic, serotonergic, and dopaminergic neurotransmitter systems (Dewey *et al*, 1990, 1992, 1993a, 1995). In the present study, we used ^{11}C -raclopride in combination with PET to study the effect of a range of PCP doses on synaptic DA. In addition, we examined the effects of increased GABAergic inhibition on this response. Finally, we used ^{11}C -cocaine to define more accurately contributions of DAT blockade to the possible DA-enhancing effects of PCP.

MATERIALS AND METHODS

Baboon PET Studies

Adult female baboons (*Papio anubis*, 14.5–18 kg) were prepared for PET scanning as detailed previously (Dewey *et al*, 1992). Animals were initially immobilized with Saffan (~10 ml) and subsequently maintained on gas anesthesia using isoflurane, nitrous oxide, and oxygen for the duration of the PET study. ^{11}C -raclopride was synthesized as previously described (specific activity, 500–900 mCi/ μmol). In all studies, dynamic PET scanning commenced simultaneously with ^{11}C -raclopride injection and was performed for 60 min in a Siemens HR+ using the following scanning protocol: 10 scans with a 1-min interval followed by 10 scans with a 5-min interval. For ^{11}C -cocaine (synthesized according to Fowler *et al*, 1989), the scanning protocol was four scans with a 0.5-min interval followed by four scans with a 1.0-min interval, four scans with a 2.0-min interval, four scans with a 5.0-min interval, and four scans with a 7.0-min interval.

Arterial blood was sampled continuously for the first 2 min using an automated device (Ole Dich, Hvidovre, Denmark) and then manually at 5.0, 10.0, 30.0, and 60.0 min postinjection. Selected plasma samples (1.0, 5.0, 10.0, and 30.0 min postinjection) were analyzed for the presence of unchanged radiotracer.

Pharmacologic Treatment Strategy

In the first series of studies, phencyclidine hydrochloride (PCP, Sigma Pharmaceuticals) was administered as an intravenous bolus injection at 0.1, 0.5, and 1.0 mg/kg 30 min prior to the second ^{11}C -raclopride scan. To potentiate whole-brain GABAergic transmission, γ -vinyl GABA (GVG, a specific suicide inhibitor of GABA amino-transferase, GABA-T; Jung *et al*, 1977) was administered 2.5 h prior to PCP administration. The inhibition of GABA-T has been shown to elevate GABA concentrations significantly in the human CNS, maximally between 2 and 4 h following parenteral administration (Palfreyman *et al*, 1981).

In the next series of studies, animals received a baseline ^{11}C -cocaine scan followed by PCP (0.5 and 1.0 mg/kg) given 30 min prior to or coadministered with the second ^{11}C -cocaine injection. For the coadministration studies, a solution of PCP was prepared prior to the second radiotracer delivery and injected into the vial containing the ^{11}C -cocaine, immediately prior to intravenous injection and subsequent scanning.

Design of Studies

All scanning protocols involved two ^{11}C -raclopride or ^{11}C -cocaine injections, while each animal remained in the tomograph. The first scan served as the baseline for the second, postchallenge scan.

We previously established the reproducibility of ^{11}C -raclopride binding as well as the effects of GVG on this binding in *papio anubis* baboons, using a test/retest experimental protocol in which each animal ($n=7$) served as its own control (Dewey *et al*, 1992). PCP was administered 30 min prior to the second ^{11}C -raclopride scan, according to a time course established by Blin *et al* (1991). These studies were performed using 0.1 mg/kg ($n=1$), 0.5 mg/kg ($n=6$), and 1.0 mg/kg ($n=3$) PCP. In a separate group of animals, GVG was administered intravenously (300 mg/kg in 4.0 ml over 3 min) immediately following completion of the first dynamic scan ($n=4$). After 2.5 h, PCP was administered 30 min prior to the second ^{11}C -raclopride scan.

Finally, we examined the time course and effects of PCP on ^{11}C -cocaine binding by administering PCP (1.0 mg/kg) either 30 min prior to the second ^{11}C -cocaine scan or simultaneously (0.5 or 1.0 mg/kg) with ^{11}C -cocaine.

Region of Interest Selection

Regions of interest (ROI) for the corpus striatum were drawn directly on the PET image to encompass the entire structure on every slice upon which it appeared and where appropriate in size for the resolution of the tomograph. This multiplanar method of ROI selection reduces differences that may arise due to movement of the animal within the

gantry during the scanning interval (Dewey *et al*, 1990). ROIs were then copied directly from the first scan to the appropriate slices of the second. Further analysis was targeted at separating the dorsal and ventral striata according to methods described by Drevets *et al* (1999). Cerebellar ROIs were drawn at the level of the vermis and included both cortical gray and white matter.

Data Analysis

Receptor availability as a function of changes in endogenous DA concentration was analyzed using a graphical technique specifically designed for reversible systems. This technique gives a linear function of the free receptor concentration known as the distribution volume (DV; Logan *et al*, 1990). The ratio of striatal to cerebellar distribution volumes (DVR) is less sensitive to noise than the individual kinetic parameters, which often have large standard errors associated with their determination. In addition, the DVR is independent of blood flow, since blood flow terms appear in the numerator and denominator and subsequently cancel out (K_1/k_2 ; Logan *et al*, 1994). The use of the DVR is also supported by the observations that the striatum and cerebellum have comparable amounts of nonspecific binding (Farde *et al*, 1989) and that our preliminary studies demonstrated that PCP did not alter the cerebellar DV in a systematic manner (either increase or decrease) in excess of the test-retest variability for this region.

We quantified ^{11}C -cocaine binding using a similar tracer kinetic modeling approach applied to differences in the availability of striatal transport proteins between test/retest animals or those pretreated with PCP. One factor that might affect estimates of binding site densities based on PET measurements is variability in the local concentration of DA; however, recent studies have demonstrated that ^{11}C -cocaine is relatively insensitive to changes in endogenous DA (Gatley *et al*, 1995), consistent with *in vitro* competition experiments with DA, suggesting that cocaine binds about 100 times more strongly at the DAT than DA (Madras *et al*, 1989). However, both ^{11}C -raclopride and ^{11}C -cocaine possess a rapid dissociation constant (Logan *et al*, 1990). This facilitates a similar fundamental kinetic modeling approach, so as with ^{11}C -raclopride, DVRs for striatum and cerebellum can be related to the ^{11}C -cocaine-binding site kinetic parameters by the ratio of striatal to cerebellar DVs.

Statistical Analysis

The statistical analysis was designed to test whether there was a difference in radiotracer binding between the dorsal and ventral tiers of the striatum and then whether this difference was a function of drug treatment prior to the second scan. Thus, differences between dorsal and ventral striata were evaluated with a one-way analysis of variance (ANOVA) across baseline scans and both baseline and challenge data were then subjected to a two-way repeated measures ANOVA with treatment group and region (dorsal vs ventral) as factors. Where appropriate, *post hoc* significance testing was performed with a Bonferroni *t*-test. Finally, to reduce the number of comparisons and to take into account the baseline condition, difference scores were

computed for the striatum, cerebellum, and the striatum to cerebellum ratio, using the formula $(\text{DVR}_{\text{Baseline Scan}} - \text{DVR}_{\text{Challenge Scan}}) / \text{DVR}_{\text{Baseline Scan}}$. The difference scores were subjected to a one-way ANOVA, with treatment group as a between-subject factor. The mean time-activity of ^{11}C -raclopride (corrected for the injected dose) was also subjected to a repeated measures ANOVA to address specific changes in radiotracer washout between animals.

RESULTS

^{11}C -Raclopride PET Scans

Immediately following administration of ^{11}C -raclopride, radioactivity accumulated in the corpus striatum bilaterally. In the cerebellum, radioactivity began to clear within the first 3 min and reached 50% of the peak within 5 min. Striatal radioactivity continued to accumulate and reached a peak value within 5 min and cleared to 50% of the peak within 25 min. In terms of the DVR, the test/retest reproducibility was determined to be 2.6% from the first to the second ^{11}C -raclopride scan (Table 1; test/retest reliability derived from $\text{DVR}_{\text{Baseline}}$ and $\text{DVR}_{\text{Challenge}}$; 3.31 ± 0.21 and 3.24 ± 0.23 , respectively; $t = 1.023$, $p = 0.325$).

^{11}C -raclopride DVR in all baseline scans was slightly, although not significantly, lower in the dorsal vs ventral striatum ($n = 18$, $\text{DVR} \pm \text{SEM} = 3.19 \pm 0.09$ vs 3.44 ± 0.1 , respectively; $F_{[1,34]} = 3.707$, $p = 0.063$). There was no significant interaction between treatment group and region, such that PCP treatment did not selectively influence ^{11}C -raclopride binding in the dorsal or ventral striatum ($F_{[1,4]} = 0.472$, $p = 0.756$), although there was a significant interaction between treatment group and scan when both regions were combined ($F_{[1,4]} = 7.979$, $p = 0.002$). Detailed analysis of this effect indicated a significant decline in ^{11}C -raclopride binding from baseline to postchallenge in animals treated with 0.5 mg/kg PCP ($\text{DVR}_{\text{Baseline}}$ and $\text{DVR}_{\text{Challenge}} = 3.52 \pm 0.09$ and 3.00 ± 0.12 , respectively; $t = 8.80$, $p < 0.001$) and 1.0 mg/kg PCP ($\text{DVR}_{\text{Baseline}}$ and $\text{DVR}_{\text{Challenge}} = 3.14 \pm 0.15$ and 2.88 ± 0.16 , respectively; $t = 3.033$, $p < 0.01$). These decreases appeared to be greater in the ventral compared to the dorsal striatum. There were no significant differences in ^{11}C -raclopride binding from baseline to challenge scans when animals were given the lowest dose of PCP (0.1 mg/kg; $\text{DVR}_{\text{Baseline}}$ and $\text{DVR}_{\text{Challenge}} = 3.65$ and 3.57 , respectively; $t = 0.546$, $p = 0.594$) or GVG prior to 0.5 mg/kg PCP ($\text{DVR}_{\text{Baseline}}$ and $\text{DVR}_{\text{Challenge}} = 3.25 \pm 0.29$ and 3.14 ± 0.21 , respectively; $t = 1.501$, $p = 0.157$). None of the doses of PCP or GVG prior to PCP altered cerebellar radioactivity, the systemic rate of labeled raclopride metabolism, or the metabolite corrected plasma input function. Figure 1 illustrates the graphical analysis applied to data from two separate animals, one given PCP alone (Figure 1a) and the other pretreated with GVG and PCP (Figure 1b).

Figures 2 and 3 demonstrate the temporal binding parameters of ^{11}C -raclopride. We observed significant differences across time points between the baseline and postchallenge scans in the group given PCP alone (Figure 2), but not in the group pretreated with GVG and then PCP (Figure 3). In primates who received only PCP, a repeated

Table 1 Percent Change in the Striatal DVR of ^{11}C -Raclopride or ^{11}C -Cocaine Produced by PCP or a Combination of PCP and the GABA-T Inhibitor, GVG

Group	Mean % change in DVR \pm SEM		
	Dorsal	Ventral	Both
^{11}C -raclopride			
Test/retest ^{11}C -raclopride ($n=4$)	-4.13 ± 3.93	-2.97 ± 1.38	-2.26 ± 3.89
GVG (300 mg/kg; $n=6$)	$7.06 \pm 1.48^\dagger$	$9.41 \pm 3.45^\dagger$	$19.01 \pm 2.2^\dagger$
PCP (0.1 mg/kg; $n=1$)	-1.07	-2.2	-2.19
PCP (0.5 mg/kg; $n=6$)	-13.49 ± 1.82	$-15.13 \pm 2.69^\dagger$	$-14.96 \pm 2.21^\dagger$
PCP (1.0 mg/kg; $n=3$)	-5.43 ± 1.8	-9.78 ± 1.07	-8.18 ± 1.12
GVG (300 mg/kg) + PCP (0.5 mg/kg; $n=4$)	-1.43 ± 5.75^i	$-3.86 \pm 3.02^\ddagger$	$-3.38 \pm 3.13^\ddagger$
^{11}C -cocaine			
Test/retest ^{11}C -cocaine ($n=3$)	1.01 ± 4.69	-1.29 ± 5.03	-0.49 ± 4.65
PCP (1.0 mg/kg) 30 min prior ($n=2$)	-10.23 ± 5.33	-9.12 ± 5.79	-9.59 ± 5.78
PCP (0.5 mg/kg) coadministered ($n=3$)	$-26.64 \pm 3.29^\S$	$-24.71 \pm 4.41^\S$	$-25.54 \pm 4.26^\S$
PCP (1.0 mg/kg) coadministered ($n=1$)	-14.10	-18.11	-13.52

† Significantly different from ^{11}C -raclopride test/retest, $p < 0.05$.
 ‡ Significantly different from ^{11}C -raclopride PCP (0.5 mg/kg), $p < 0.05$.
 i Significantly different from ^{11}C -raclopride PCP (0.5 mg/kg), $p < 0.01$.
 § Significantly different from ^{11}C -cocaine test/retest, $p < 0.05$.

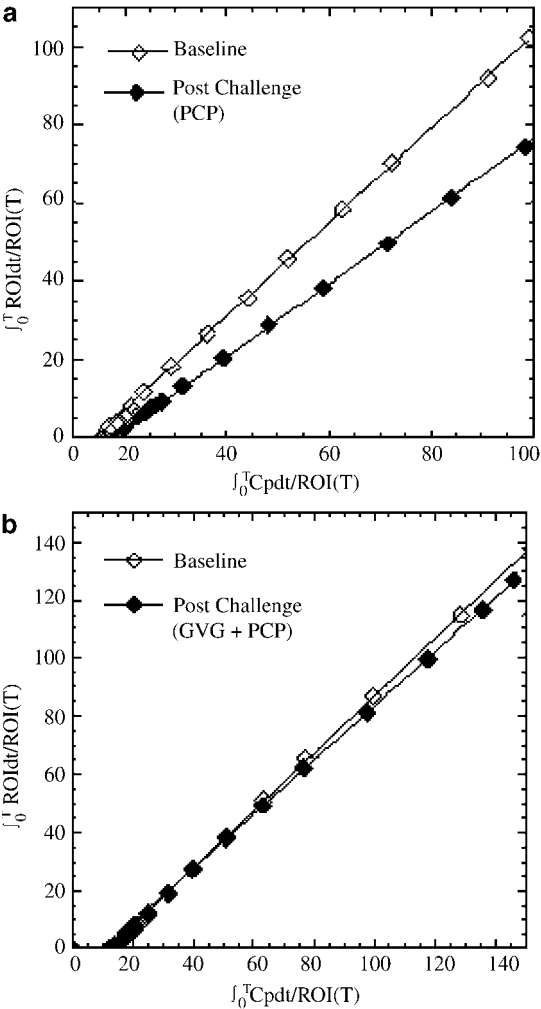


Figure 1 Graphical analysis of the ^{11}C -raclopride time-activity data from striatal ROIs in (a) one primate given a 0.5 mg/kg PCP challenge and (b) a primate pretreated with GVG (300 mg/kg) followed by the same PCP challenge. ROI(T) refers to radioactivity in the striatum at time T . $C_p(t)$ is the plasma radioactivity concentration corrected for metabolites.

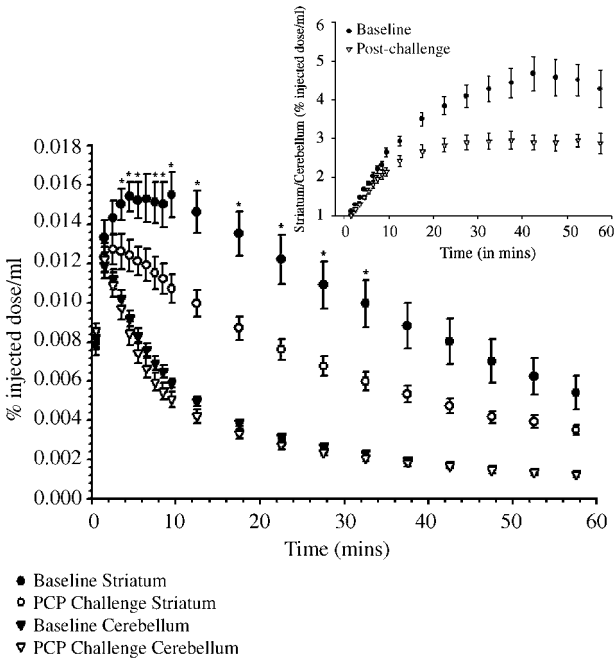


Figure 2 Time-activity data from striatum and cerebellum ROIs from five PET studies where primates were given a PCP challenge (0.5 mg/kg) prior to the second ^{11}C -raclopride scan. All points on the graph are corrected for the presence of labeled raclopride metabolites, and represent mean \pm standard error for all three animals. Univariate repeated measures ANOVA indicates a significant difference between baseline and post-challenge (PCP alone) conditions at a given time, T (* $p < 0.05$).

measures ANOVA indicated that significant differences between baseline and postchallenge ^{11}C -raclopride scans correlated with times of peak radioactivity (beginning 4.5 min after injection, $F = 20.79$, $p = 0.049$ –27.5 min post-injection, $F = 18.48$, $p = 0.0501$). Further, only one baseline point in the GVG + PCP time-activity curve (Figure 3) appeared significantly different from a postchallenge point (occurring 7.5 min postinjection, $F = 8.50$, $p = 0.0427$). Separate analysis of striatal and cerebellar values indicated

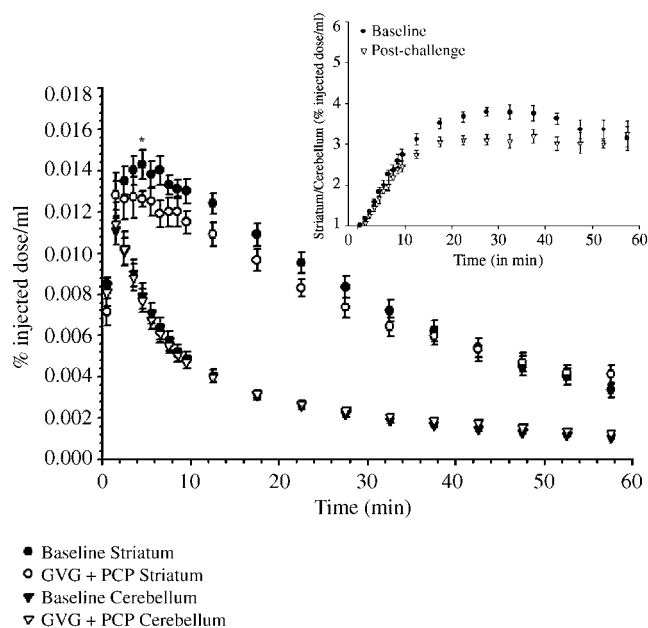


Figure 3 Time-activity data from striatum and cerebellum ROIs from four PET studies where primates were pretreated with GVG (300 mg/kg) prior to a PCP challenge (0.5 mg/kg) before the second ^{11}C -raclopride scan. All points are corrected for the presence of labeled raclopride metabolites, and represent mean \pm standard error for all five animals. Univariate repeated measures ANOVA indicates a significant difference between baseline and postchallenge (GVG + PCP) conditions at a given time, T (* $p < 0.05$).

significant between-group differences in striatal activity from baseline to postchallenge ($F = 45.60$, $p < 0.0001$), but not in cerebellar activity ($F = 2.71$, $p = 0.108$).

^{11}C -Cocaine PET Scans

There was a marked accumulation of ^{11}C -cocaine in striatal regions, whereas levels of radioactivity in the cerebellum were much lower in all groups. Analysis of radioactivity-corrected time-activity curves indicates that the striatal accumulation of ^{11}C -cocaine peaked around 5.0 min, and rapidly declined. The test/retest reliability of ^{11}C -cocaine is given in Table 1. Regional analysis of all baseline ^{11}C -cocaine specific binding in the dorsal vs ventral striata indicated that there were no regional effects ($n = 8$, $\text{DVR} \pm \text{SEM} = 2.13 \pm 0.16$ vs 2.3 ± 0.2 , respectively; $F_{[1,14]} = 0.679$, $p = 0.424$). However, there appears to be a significant interaction between treatment group and DVR when the regions were combined, such that a significant reduction in ^{11}C -cocaine binding depended on the PCP treatment regimen ($F_{[3,4]} = 9.94$, $p = 0.025$). Detailed analyses of interactions between PCP and DAT sites indicate a significant inhibition of ^{11}C -cocaine binding only when coadministered with 0.5 mg/kg PCP ($\text{DVR}_{\text{Baseline}}$ and $\text{DVR}_{\text{Challenge}} = 2.55 \pm 0.23$ and 1.90 ± 0.28 , respectively; $t = 6.830$, $p = 0.002$). At 1.0 mg/kg given 30 min prior to ^{11}C -cocaine or coadministered, there were no significant effects (30 min pretreat: $\text{DVR}_{\text{Baseline}}$ and $\text{DVR}_{\text{Challenge}} = 1.83 \pm 0.10$ and 1.65 ± 0.20 , respectively; $t = 1.839$, $p = 0.140$ and coadministered: $\text{DVR}_{\text{Baseline}}$ and $\text{DVR}_{\text{Challenge}} = 2.44$ and 2.11 , respectively).

DISCUSSION

In the present study, we used PET to explore the hypothesis that targets outside the NMDA receptor complex contribute to PCP-induced decreases in ^{11}C -raclopride binding. First, we provide evidence that GABAergic systems are involved in this response, since it can be attenuated with GVG pretreatment. Second, our data demonstrate a potent interaction between PCP and the ^{11}C -cocaine-binding site in the living primate, which may contribute to decreases in striatal ^{11}C -raclopride binding by preventing reuptake of synaptic DA, in addition to blocking norepinephrine and serotonin reuptake. Together, these experiments contribute to a growing body of literature suggesting that NMDA antagonist mediated changes in striatal DA may be partially mediated by GABAergic systems, and also by blockade of monoamine transporters.

PET studies in humans demonstrated that ketamine significantly reduced the binding of ^{11}C -raclopride, consistent with increased competition from synaptic DA (Breier *et al*, 1998; Smith *et al*, 1998; Vollenweider *et al*, 2000). However, recent microdialysis findings suggest that PCP may not increase striatal ECF DA in nonhuman primates (Adams *et al*, 2002), and that ketamine may not alter ^{11}C -raclopride binding in humans (Kegeles *et al*, 2002). Studies by Tsukada *et al* (2000) may present some resolution to this problem. They combined microdialysis and PET measures in conscious monkeys to demonstrate that ketamine decreased ^{11}C -raclopride binding, but did not affect extracellular DA. Further, Tsukada *et al* (2001) attribute ketamine's lack of effect on extracellular systems to an increased efficacy of DAT sites, while our data suggest that PCP specifically, is a more potent blocker of these proteins than was previously thought. Thus, microdialysis measures of increases in extracellular DA in response to PCP might reflect spillover secondary to monoamine transporter blockade at higher doses (Schiffer *et al*, 2001a), while lower doses of PCP may be absent from this effect. In fact, recent findings using MK-801 have provided evidence that this drug increases the availability of DAT sites (Nakano *et al*, 1998; Page *et al*, 2000; Schiffer *et al*, 2001b; Kagawa *et al*, 2002). Taken together, it appears that individual NMDA antagonists might produce compound-specific effects on striatal dopaminergic systems as a function of their affinity for targets outside the NMDA receptor complex. In agreement with this, recent *in vitro* data also suggest that ketamine and to a lesser extent PCP, compete directly with ^3H -raclopride binding at the high affinity D_2 receptor site (Kapur and Seeman, 2001, 2002). Therefore, it appears that there may be a multitude of mechanisms by which PCP increases extracellular DA or alters ^{11}C -raclopride binding.

If NMDA receptor antagonism diminishes GABA neuron firing (Hondo *et al*, 1995), then systemically increasing GABAergic inhibition (via GABA_A or GABA_B receptor agonists) should have little consequence. In fact, our previous *in vivo* microdialysis results in freely moving animals demonstrated that increased GABAergic activity produced a preferential inhibition of cortical over subcortical DA release (Schiffer *et al*, 2001a). Similarly, local application of specific GABA receptor agonists (muscimol, baclofen, or bicuculline) in cortical (Yonezawa *et al*, 1998) or subcortical (Westerink *et al*, 1996) areas produced a

similar inhibition of the same system. Consistent with these studies, electrophysiological measures in the hippocampus suggest that disinhibition of GABAergic regulatory processes by NMDA antagonists may be a primary mechanism associated with NMDA antagonist-induced stimulation (Grunze *et al*, 1996). Combined with behavioral evidence indicating that GVG and other GABA agonists can diminish some of the stereotypies associated with PCP (Seiler and Grauffel, 1992), it appears that at least some, if not all, of the GABAergic inhibitory control is spared from the effects of NMDA receptor antagonism.

It is possible that these 'spared' GABAergic effects are also a function of the dose of PCP, and may be related to its additional molecular targets. We demonstrate a significant, reproducible decrease in ^{11}C -cocaine binding, consistent with direct competition for the DAT and perhaps a shared mechanism. Interestingly, this effect was less pronounced at the highest and lowest doses of PCP. This provides one explanation for previous findings in which doses below those used here did not increase striatal extracellular DA, nor did they markedly influence nomifensine-induced DA overflow (Adams *et al*, 2002). The present PET studies suggest an inverted U-shaped dose-response curve for both inhibiting ^{11}C -raclopride and ^{11}C -cocaine binding, where 0.5 mg/kg PCP produced a greater inhibition of both radiotracers than 1.0 mg/kg. Thus, doses of 1.0 mg/kg PCP may exceed the psychotomimetic and addictive effects of PCP, and our results at these doses represent those that might occur during anesthesia or during preanesthetic ataxic states associated with PCP. Consistent with this latter hypothesis, we did not have to alter our anesthesia at any time following PCP administration and there were no changes in vital signs following its administration.

Direct competition with transporter proteins does not completely explain the present finding that increasing GABAergic tone attenuates PCP-induced decreases in ^{11}C -raclopride binding. For example, in the present study, while GVG attenuated PCP-induced decreases in ^{11}C -raclopride binding, it totally abolished cocaine-induced decreases in ^{11}C -raclopride binding (Dewey *et al*, 1998). However, effects reported with cocaine may also involve other neurotransmitter systems including serotonin, acetylcholine, and norepinephrine. Consistent with this notion of multi-transmitter effects, a component of the PCP-induced effects may be related to NMDA receptor modulation. In fact, comparisons of PCP and MK-801, which have little or no affinity for the DAT (Maurice *et al*, 1991), indicate that both drugs possess rewarding and reinforcing properties (Willins, 1993), even though there is little support for an excitatory effect of MK-801 on DA activity (Mele *et al*, 1998). Further, if the present reductions in ^{11}C -raclopride binding were due to a direct competition for D_2 receptor sites (Kapur and Seeman, 2001, 2002), then it is unlikely that *a priori* increases in GABAergic tone would influence this effect. Thus, it appears that PCP-induced decreases in ^{11}C -raclopride binding are related to a combination of direct stimulation from glutamatergic spillover (Adams and Moghaddam, 1998) and DAT blockade itself (Schiffer *et al*, 2001a) and that like cocaine, these decreases are susceptible to partial modulation by GABAergic systems.

Finally, these studies continue to support our premise that moderate affinity radiotracers provide a dynamic

measure of competition from synaptic DA, which appear sensitive to pharmacologic manipulations of dopaminergic, serotonergic, GABAergic, cholinergic, and now glutamatergic systems (Dewey *et al*, 1993b, 1995; Schloesser *et al*, 1996). PET therefore provides a unique opportunity to observe and compare the cascade of neurochemical events produced by drugs that appear to have similar initial targets. These data may have implications for the development of unique therapeutic strategies targeted at specific neurotransmitter systems currently implicated in schizophrenia and addiction.

ACKNOWLEDGEMENTS

We gratefully acknowledge J Fowler, P Vaska, D Alexoff, D Schlyer, P King, P Carter, N Pappas, M Gerasimov, C Shea, V Garza, and R Ferrieri. Hoescht Marion Roussel and ChiroTech supplied the GVG. This research was carried out at Brookhaven National Laboratory under contract with the US Department of Energy Office of Biological and Environmental Research (USDOE/OBER DE-AC02-98CH10886), and partially funded by NIH awards to WKS (F31-DA15874) and SLD (DA015041).

REFERENCES

- Adams B, Moghaddam B (1998). Corticolimbic dopamine neurotransmission is temporally dissociated from the cognitive and locomotor effects of phencyclidine. *J Neurosci* 18: 5545–5554.
- Adams BW, Bradberry CW, Moghaddam B (2002). NMDA antagonist effects on striatal dopamine release: microdialysis studies in awake monkeys. *Synapse* 43: 12–18.
- Blin J, Denis A, Yamaguchi T, Crouzel C, MacKenzie ET, Baron JC (1991). PET studies of [18F]methyl-MK-801, a potential NMDA receptor complex radioligand. *Neurosci Lett* 121: 183–186.
- Breier A, Adler CM, Weisenfeld N, Su TP, Elman I, Picken L *et al* (1998). Effects of NMDA antagonism on striatal dopamine release in healthy subjects: application of a novel PET approach. *Synapse* 29: 142–147.
- Carlezon WA, Wise RA (1996). Rewarding actions of phencyclidine and related drugs in nucleus accumbens shell and frontal cortex. *J Neurosci* 16: 3112–3122.
- Dewey SL, Brodie JD, Fowler JS, MacGregor RR, Schlyer DJ, King PT *et al* (1990). Positron emission tomography (PET) studies of dopaminergic/cholinergic interactions in the baboon brain. *Synapse* 6: 321–327.
- Dewey SL, Morgan AE, Ashby Jr CR, Horan B, Kushner SA, Logan J *et al* (1998). A novel strategy for the treatment of cocaine addiction. *Synapse* 30: 119–129.
- Dewey SL, Smith GS, Logan J, Alexoff D, Ding YS, King P *et al* (1995). Serotonergic modulation of striatal dopamine measured with positron emission tomography (PET) and *in vivo* microdialysis. *J Neurosci* 15: 821–829.
- Dewey SL, Smith GS, Logan J, Brodie JD (1993a). Modulation of central cholinergic activity by GABA and serotonin: PET studies with 11C-benzotropine in primates. *Neuropsychopharmacology* 8: 371–376.
- Dewey SL, Smith GS, Logan J, Brodie JD, Simkowitz P, MacGregor RR *et al* (1993b). Effects of central cholinergic blockade on striatal dopamine release measured with positron emission tomography in normal human subjects. *Proc Natl Acad Sci USA* 90: 11816–11820.
- Dewey SL, Smith GS, Logan J, Brodie JD, Yu DW, Ferrieri RA *et al* (1992). GABAergic inhibition of endogenous dopamine release

- measured *in vivo* with ¹¹C-raclopride and positron emission tomography. *J Neurosci* 12: 3773–3780.
- Drevets WC, Price JC, Kupfer DJ, Kinahan PE, Lopresti B, Holt D *et al* (1999). PET measures of amphetamine-induced dopamine release in ventral vs dorsal striatum. *Neuropsychopharmacology* 21: 694–709.
- Farde L, Eriksson L, Blomquist G, Halldin C (1989). Kinetic analysis of central [¹¹C]raclopride binding to D₂-dopamine receptors studied by PET—a comparison to the equilibrium analysis. *J Cereb Blood Flow Metab* 9: 696–708.
- Fowler JS, Volkow ND, Wolf AP, Dewey SL, Schlyer DJ, Macgregor RR *et al* (1989). Mapping cocaine binding sites in human and baboon brain *in vivo*. *Synapse* 4: 371–377.
- Gatley SJ, Volkow ND, Fowler JS, Dewey SL, Logan J (1995). Sensitivity of striatal [¹¹C]cocaine binding to decreases in synaptic dopamine. *Synapse* 20: 137–144.
- Grunze HC, Rainnie DG, Hasselmo ME, Barkai E, Hearn EF, McCarley RW *et al* (1996). NMDA-dependent modulation of CA1 local circuit inhibition. *J Neurosci* 16: 2034–2043.
- Hondo H, Nakahara T, Nakamura K, Hirano M, Uchimura H, Tashiro N (1995). The effect of phencyclidine on the basal and high potassium evoked extracellular GABA levels in the striatum of freely moving rats: an *in vivo* microdialysis study. *Brain Res* 671: 54–62.
- Johnson KM, Snell LD (1985). Effects of phencyclidine (PCP)-like drugs on turning behavior, 3H-dopamine uptake, and 3H-PCP binding. *Pharmacol Biochem Behav* 22: 731–735.
- Jung MJ, Lippert B, Metcalf BW, Bohlen P, Schechter PJ (1977). gamma-Vinyl GABA (4-amino-hex-5-enoic acid), a new selective irreversible inhibitor of GABA-T: effects on brain GABA metabolism in mice. *J Neurochem* 29: 797–802.
- Kagawa S, Nakano T, Inoue O, Nishimura T (2002). Effect of glutamatergic systems on *in vivo* binding of [(125)I]beta-CIT in the brain of a rat model of Parkinson's disease. *Synapse* 46: 38–44.
- Kapur S, Seeman P (2001). Ketamine has equal affinity for NMDA receptors and the high-affinity state of the dopamine D₂ receptor. *Biol Psychiatry* 49: 954–957.
- Kapur S, Seeman P (2002). NMDA receptor antagonists ketamine and PCP have direct effects on the dopamine D(2) and serotonin 5-HT(2) receptors—implications for models of schizophrenia. *Mol Psychiatry* 7: 837–844.
- Kegeles LS, Martinez D, Kochan LD, Hwang DR, Huang Y, Mawlawi O *et al* (2002). NMDA antagonist effects on striatal dopamine release: positron emission tomography studies in humans. *Synapse* 43: 19–29.
- Li Q, Clark S, Lewis DV, Wilson WA (2002). NMDA receptor antagonists disinhibit rat posterior cingulate and retrosplenial cortices: a potential mechanism of neurotoxicity. *J Neurosci* 22: 3070–3080.
- Logan J, Fowler JS, Volkow ND, Wolf AP, Dewey SL, Schlyer DJ *et al* (1990). Graphical analysis of reversible radioligand binding from time-activity measurements applied to [¹¹C-methyl]-(-)-cocaine PET studies in human subjects. *J Cereb Blood Flow Metab* 10: 740–747.
- Logan J, Volkow ND, Fowler JS, Wang GJ, Dewey SL, MacGregor R *et al* (1994). Effects of blood flow on [¹¹C]raclopride binding in the brain: model simulations and kinetic analysis of PET data. *J Cereb Blood Flow Metab* 14: 995–1010.
- Madras BK, Fahey MA, Bergman J, Canfield DR, Spealman RD (1989). Effects of cocaine and related drugs in nonhuman primates. I. [³H]cocaine binding sites in caudate-putamen. *J Pharmacol Exp Ther* 251: 131–141.
- Maurice T, Vignon J, Kamenka JM, Chicheportiche R (1991). Differential interaction of phencyclidine-like drugs with the dopamine uptake complex *in vivo*. *J Neurochem* 56: 553–559.
- Mele A, Wozniak KM, Hall FS, Pert A (1998). The role of striatal dopaminergic mechanisms in rotational behavior induced by phencyclidine and phencyclidine-like drugs. *Psychopharmacology* 135: 107–118.
- Nakano T, Takatoku K, Matoba Y, Iwamoto B, Nishiura M, Inoue O *et al* (1998). Enhancement of *in vivo* binding of [¹²³I]beta-CIT by MK-801 in rat brain. *Synapse* 30: 402–408.
- Olney JW, Farber NB (1995). Glutamate receptor dysfunction and schizophrenia. *Arch Gen Psychiatry* 52: 998–1007.
- Page G, Peeters M, Maloteaux JM, Hermans E (2000). Increased dopamine uptake in striatal synaptosomes after treatment of rats with amantadine. *Eur J Pharmacol* 403: 75–80.
- Palfreyman MG, Schechter PJ, Buckett WR, Tell GP, Koch-Weser J (1981). The pharmacology of GABA-transaminase inhibitors. *Biochem Pharmacol* 30: 817–824.
- Rodefer JS, Carroll ME (1999). Concurrent progressive-ratio schedules to compare reinforcing effectiveness of different phencyclidine (PCP) concentrations in rhesus monkeys. *Psychopharmacology (Berl)* 144: 163–174.
- Schiffer WK, Gerasimov M, Hofmann L, Marsteller D, Ashby CR, Brodie JD *et al* (2001a). Gamma vinyl-GABA differentially modulates NMDA antagonist-induced increases in mesocortical vs mesolimbic DA transmission. *Neuropsychopharmacology* 25: 704–712.
- Schiffer WK, Gerasimov MR, Fowler JS, Volkow ND, Gifford A, Ashby CR *et al* (2001b). MK-801 increases dopamine transporter (DAT) availability: a positron emission tomography (PET) investigation in non-human primates. *J Nucl Med* 42: 56.
- Schloesser R, Simkowitz P, Bartlett EJ, Wolkin A, Smith GS, Dewey SL *et al* (1996). The study of neurotransmitter interactions using positron emission tomography and functional coupling. *Clin Neuropharmacol* 19: 371–389.
- Seiler N, Grauffel C (1992). Antagonism of phencyclidine-induced hyperactivity in mice by elevated brain GABA concentrations. *Pharmacol Biochem Behav* 41: 603–606.
- Smith GS, Schloesser R, Brodie JD, Dewey SL, Logan J, Vitkun SA *et al* (1998). Glutamate modulation of dopamine measured *in vivo* with positron emission tomography (PET) and ¹¹C-raclopride in normal human subjects. *Neuropsychopharmacology* 18: 18–25.
- Tsukada H, Harada N, Nishiyama S, Ohba H, Sato K, Fukumoto D *et al* (2000). Ketamine decreased striatal [(11)C]raclopride binding with no alterations in static dopamine concentrations in the striatal extracellular fluid in the monkey brain: multi-parametric PET studies combined with microdialysis analysis. *Synapse* 37: 95–103.
- Tsukada H, Nishiyama S, Kakiuchi T, Ohba H, Sato K, Harada N (2001). Ketamine alters the availability of striatal dopamine transporter as measured by [(11)C]beta-CFT and [(11)C]beta-CIT-FE in the monkey brain. *Synapse* 42: 273–280.
- Volkow ND, Wang GJ, Fowler JS, Logan J, Gatley SJ, Wong C *et al* (1999). Reinforcing effects of psychostimulants in humans are associated with increases in brain dopamine and occupancy of D(2) receptors. *J Pharmacol Exp Ther* 291: 409–415.
- Vollenweider FX, Vontobel P, Oye I, Hell D, Leenders KL (2000). Effects of (S)-ketamine on striatal dopamine: a [¹¹C]raclopride PET study of a model psychosis in humans. *J Psychiatr Res* 34: 35–43.
- Westerink BH, Kwint HF, deVries JB (1996). The pharmacology of mesolimbic dopamine neurons: a dual-probe microdialysis study in the ventral tegmental area and nucleus accumbens of the rat brain. *J Neurosci* 16: 2605–2611.
- Willins D (1993). The role of dopamine and AMPA/kainate receptors in the nucleus accumbens in the hypermotility response to MK-801. *Pharmacol Biochem Behav* 46: 881–887.
- Yonezawa Y, Kuroki T, Kawahara T, Tashiro N, Uchimura H (1998). Involvement of gamma-aminobutyric acid neurotransmission in phencyclidine-induced dopamine release in the medial prefrontal cortex. *Eur J Pharmacol* 341: 45–56.